This factsheet describes baiting systems used for managing termites in Australia. Termite baiting concepts, termite aggregation in baiting stations and toxicant application are explored with respect to feeding habits and behaviour of some termites that damage timber-in-service in Australia. Information is also given about the economic impact of termites, the history and development of termite baiting systems and commercial termite baiting systems.

Until recently, subterranean termite management in Australia relied on chemically-treated soil barriers using the highly persistent cyclodiene (organochlorine) insecticides (aldrin, dieldrin, chlordane and heptachlor). These chemicals were de-registered and withdrawn from use from 30 June 1995. Well before this date, alternative strategies for subterranean termite management in buildings and structures were being developed. One of these uses so-called ‘termite baiting systems’.

Economic importance

Australia’s diverse termite fauna includes 258 described, and at least 90 undescribed termite species from about 30 genera in five families (Mastotermitidae, Termopsidae, Kalotermitidae, Rhinotermitidae and Termitidae). Termites are grouped broadly into three categories: dampwood, drywood and subterranean termites.

Most species of termites that damage timber-in-service in Australia are subterranean termites. Mastotermes darwiniensis and Coptotermes spp. are the most destructive. Coptotermes acinaciformis (Figure 1) is responsible for more economic loss than all the other Australian species combined. This is because it has an extensive range, causes severe damage and can survive in cities and large towns. Some species of Schedorhinotermes and Nasutitermes are also important economically. Nasutitermes exitiosus is particularly widespread in the cool temperate regions of the southern, mainland states.

The annual cost of termite damage to buildings in Australia is estimated to exceed $100 million, while the cost of imported chemicals for termite management may exceed $20 million annually. Whatever the real figures, termites can cause significant damage with devastating financial and social implications for building owners.

Feeding habits and behaviour

Cellulose is the basic food requirement of all termites, and all types of plant material can be damaged. Most termite species eat grass and other surface vegetation and have an important role in maintaining soil fertility and aeration. Other termite species infest timber, particularly in an early state of decay caused by wood-rotting fungi.

Several timber species have natural properties that make the heartwood resistant to termite damage. Most termite species which infest timber can damage the sapwood of softwoods, for example hoop, slash and radiata pines. However, wood-feeding Nasutitermes spp. only damage hardwoods.

Figure 1. Coptotermes acinaciformis soldier.
Subterranean termites forage for food using covered runways (galleries) extending from the central nest to food sources above (Figure 2) or below ground. The gallery system of a single colony may exploit food sources over as much as one hectare, and individual galleries may be up to 50 m in long. In the case of the giant northern termite *M. darwiniensis*, individual galleries may extend as far as 200 m. Usually, only workers and soldiers visit the feeding sites, but nymphs are sometimes present. Other members of the colony generally remain in the nest. Foraging activity is seasonal, and may slow down or even stop during winter. Most termites need a plentiful supply of water, part of which they obtain from damp soil. Foraging termites aggregate in warm, moist areas containing susceptible timber, although exploratory foraging is thought to be at random.

Termite baiting concepts

Although termites are not strictly attracted, enticed or lured to most baits, as in a conventional baiting system, the principle is to have a susceptible substance in an aggregation device ('bait station') on which the termites aggregate and continue to feed once they have found the bait station. Bait stations can be placed in-ground or above-ground and placing them in areas conducive to termite activity (directed placement) improves the chance of contact with foraging termites. A bait toxicant in timber or a cellulose matrix can be placed in the station, or the colony may be destroyed indirectly by dusting aggregated termites.

Some bait toxicants have the potential to eliminate the colony while others only suppress the colony. Importantly, however, both methods reduce further, potential damage to timbers.

Termite baiting is most beneficial when used as part of an integrated pest management strategy. Colony elimination or suppression should be followed by hazard reduction and regular inspection. Monitoring should continue because only a small amount of toxin is used and will only affect a single colony. This does not prevent foraging by other termite colonies that may be present in the foraging range of a timber structure.

Termite aggregation

Several techniques have been developed to aggregate termites in stations prior to treatment in in-ground and above-ground situations. Care should be taken when inspecting stations because termites, especially *Schedorhinotermes* spp., tend to retreat from, and avoid, disturbed areas.

In-ground aggregation

Termites appear to forage randomly through the soil, but can quickly recruit large numbers to a new food source. They are thought to regulate the number of foragers at a food source directly according to the mass of susceptible timber available. This behaviour
has implications with regard to aggregating large numbers of termites. Large timbers buried in the ground are likely to be found more readily and will amass larger numbers of termites than small blocks of wood. Aggregation devices include the termite bait box (developed from a CSIRO design) and plastic conduits.

The termite bait box

There are few critical dimensions or materials. Typically, boxes are about 200x300x500 mm and constructed of polystyrene (e.g. broccoli box, Figure 3), polyethylene (e.g. worm farm box) or untreated timber, and buried in soil to about 100 mm. The lid should seal tightly, to prevent ants from entering and the contents from drying. Holes in the base of the box permit termite entry. The inside can be inspected through a small window, covered with clear plastic, in the top or end of the box. A sheet of white tissue or blotting paper against the inside of the window may be used to detect termites (because termites will eat and stain the paper) without disturbing them. Wetted corrugated cardboard and thin strips of susceptible wood are laid alternately as the bait substrate. These are easily removed to obtain the termites.

Figure 3. The termite bait box.

Plastic conduit devices

Again, there are few critical dimensions or materials. Various do-it-yourself plastic-conduit devices have been used. These include plastic conduit (25 mm diameter), perforated with small holes (4 mm diameter) drilled every 100 mm, with wetted corrugated cardboard inserted. The device is buried (100—150 mm depth) in wetted soil diagonally across the under-house area, around the outside of the building, or near active galleries. Plastic T pieces, inserted about every metre, are fitted with vertical tubing (200 mm). A small plastic box (100x130x200 mm) is attached to each of these and filled with layers of wetted corrugated cardboard to aggregate termites.

‘Stud’ size pine timber (35x70x2400 mm) with a slotted PVC stormwater pipe (500 mm long x 90 mm diameter, containing timber) positioned in the ground vertically along the stud (Figure 4) has been used successfully. A removable cap aids inspection.

Figure 4. Termite aggregation station with slotted, PVC stormwater pipe and timber before installation.

Figure 5. Installing a termite aggregation station with plastic T piece and timber.
Alternatively, a plastic T piece (90 mm diameter) with a cap can be positioned along the stud (Figure 5). PVC stormwater pipe (500x90 mm diameter, containing timber) can be attached to the T piece, as necessary.

**Above-ground aggregation**

Stations placed in direct contact with infested timbers above-ground enables foraging termites to find the bait easily. However, access to suitable sites may be difficult, especially in association with brick-veneer, slab-on-ground construction.

**Trees and stumps**

Some termite species build nests in large trees. These trees appear unaffected and although the whole trunk may be hollow, the only external signs may be hollow, broken branches. Termites can be detected by drilling the tree. All trees, especially large trees within 50 m of a building should be checked. Using a long auger bit (20 mm diameter), several holes are bored towards the centre of the tree trunk, approximately 1 m from the ground. If termites are present (or have been active previously) the trunk centre will be hollow, or filled with muddy material (‘mudgut’). The auger will suddenly penetrate easily as it reaches the hollowed centre.

There are a number of ways to test for termites in the tree. First, the auger should be examined when it is withdrawn from the tree. If termites are not found on the auger bit, insert a length of dowel into each bore hole. The diameter of the dowel should be slightly smaller than that of the hole, as the sap causes the dowel to swell. Allow at least 150 mm of the dowel to protrude to serve as a handle. The dowel should be withdrawn after one to two weeks and checked for signs of termites, which include chewed areas or brown, faecal spotting. Economically important species of termites found in trees should be treated.

Rectangular plastic containers (e.g. lunch boxes, 70x130x200 mm) are used successfully as aggregation stations in trees and timber house stumps. A 19 mm hole is drilled into each of the two longest sides of each container and a 300 mm length of 19 mm diameter plastic conduit attached through the holes. Four 10 mm diameter holes are drilled in the conduit to allow the termites to enter the box. Pieces of wetted single-backed corrugated cardboard and thin strips of susceptible wood are layered alternately as bait, and the box covered with a plastic lid. A piece of single-backed corrugated cardboard (45x350 mm) is placed in the conduit and one end plugged with a rubber stopper. The box is then connected to the colony by inserting the plastic conduit into a hole (20 mm diameter) drilled in the side of the tree or infested stump. Termite entry is verified by inspection.

**Placing baits**

In Australia, the main problem with baiting techniques for *Coptotermes* spp. has been the inconsistency of termites locating and accepting the baits.

Directed placement of in-ground stations should be in dark, moist, quiet places (especially where termite activity has been confirmed), rather than at regular intervals around a structure. Primary areas for directed placement include areas around drains, areas directly adjacent to ‘wet areas’ (bathroom, laundry and kitchen) where plumbing may enter the structure, garden beds adjacent to the structure and around trees or stumps.

Competing termite food sources cannot usually be removed, and food choices are always available to the termites being baited. In some cases, baits have been pre-decayed with a suitable fungus to improve its acceptability to termites. The fungus used varies with the termite species. Similarly, nitrogen additives, principally urea and selected amino acids, have been used to increase the feeding on baits by *Coptotermes* spp.

In above-ground stations, baits should be placed in contact with infested timbers or active termite galleries. Electronic devices to detect termite movement have been developed. One such a device is Termatrac (www.termatrac.com).

**Applying toxicant**

The station may contain a bait toxicant or ‘bait-and-switch’ techniques may be employed by replacing the initial bait with one laced with a toxicant. Alternatively, the toxin may be applied topically, usually to several thousand termites, as a dust. Whichever strategy is employed, success depends on the aggregated termites taking a slow-acting toxin back to the nest which eventually kills or suppresses the colony. The toxin-affected individuals must move away from the bait site because accumulation of dead termites will repel other individuals from approaching the bait. Ideally, the toxicant should
be non-repellent to termites, or at least masked by other agents to prevent avoidance behaviour by the foraging termites. The intent is to starve the termites by killing the gut protozoa or bacteria, or to starve the colony by upsetting the social system or to otherwise interfere with the colony.

**Bait toxicant**

Several chitin-synthesis inhibitors (for example, chlorfluazuron, flufenoxuron, hexaflumuron, noviflumuron and triflumuron) and metabolic inhibitors (for example, hydramethylnon and sulfuramid) have delayed activity against some termites. As a bait toxicant, they can be used to manage foraging populations of subterranean termite colonies, reducing the potential damage.

**Dusting**

Termites may be aggregated and dusted using a hand blower. Alternatively the aggregated termites may be removed, dusted and then returned to the site of activity. Removal and dusting should be done quickly to avoid stressing the termites and to prevent others from sealing galleries as they retreat. The removed termites are sorted into a large, flat container by tapping the bait matrix. The termites are then transferred to a jar or plastic bag and gently rolled in the dust. The dusted termites are then released back into the station. The lid of the station is left off until most of the termites have entered their galleries and left the station. With the ‘Trojan Termite Technique’, termites are brought from another colony, dusted and released into a station containing aggregated termites. The toxin, in theory, is transferred between termites and through the colony.

Arsenic trioxide is commonly used as a ‘termite dust’. Arsenic trioxide is highly toxic to humans. For the method to be effective and safe, operator skill is required in judging both the quantities and placement of dust and in handling the toxin and the termites. The treatment must be carried out only by an experienced licensed operator.

Fipronil, permethrin and triflumuron are also registered as dust applications. Experimental biological control has been successful using insect pathogenic bacterial spores, the fungus *Metarhizium anisopliae* and roundworms (nematodes) *Heterorhabdis* spp. and *Steinernema* spp. Commercial products are not yet on the Australian market.

Because each termite colony generally forages in its own gallery system, isolated from other termite colonies and organisms, it is considered that treating a colony with bait toxicants will not lead to widespread contamination of the soil. It should, therefore, not cause damage to other termite colonies, or other soil-dwelling organisms, of importance in woodland and urban ecosystems.

**Commercial termite baiting systems**

Several manufacturers have bait toxicants registered with the Australian Pesticides and Veterinary Medicines Authority (APVMA, website: [www.apvma.gov.au](http://www.apvma.gov.au)) Details of these are available from the manufacturer’s websites.

**Conclusion**

Termite-baiting systems are largely in a developmental or refinement phase in Australia. They are most beneficial when used as part of an integrated pest management strategy, and are not the complete answer to termite management. Colony elimination or suppression should be followed by hazard reduction and regular inspection. A better understanding of termite foraging habits and behaviour will enhance bait-station design and placement for termite aggregation and bait-toxicant application.

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**Source**


**More information**

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